

Distribution of Pilus island and antibiotic resistance genes in *Streptococcus agalactiae* obtained from vagina of pregnant women in Yazd, Iran

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ABSTRACT

Background and Objectives: Due to the important role of *Streptococcus agalactiae*, Group B streptococci (GBS), in production of invasive disease in neonates, investigation regarding the pathogenicity and antibiotic resistance factors is necessary in selecting the appropriate therapeutic agents. Beside capsule, the pilus has been currently recognized as an important factor in enhancing the pathogenicity of GBS. Resistance of GBS to selected antibiotics is noticeably increasing which is mainly due to the anomalous use of these drugs for treatment. The aim of this study was to determine the prevalence of pili genes followed by antibiotic susceptibility of GBS, previously serotyped, isolated from pregnant women in the city of Yazd, Iran.

Materials and Methods: Fifty seven GBS from pregnant women were subjected to multiplex PCR for determination of PI-1, PI-2a and PI-2b pilus-islands and simultaneously, the phenotype of antibiotic resistance to penicillin, tetracycline, erythromycin, clindamycin, gentamycin and levofloxacin was determined. Antibiotic resistance genes (*ermA*, *ermB*, *mefA*, *tetM*, *int-Tn*) were further diagnosed using PCR and multiplex PCR.

Results: PI-1+PI-2a with 71.9%; followed by PI-2a (21.1%) and PI-2b (7%) were observed. PI-1+PI-2a in serotype III was (73.2%), serotype II, Ia, Ib and V were 12.2%, 9.8%, 2.4% and 2.4% respectively. GBS penicillin sensitive was 89.5% and 96.5% resistance to tetracycline. The frequency of resistance genes were as follows: *tetM* (93%), *ermA* (33.3%), *ermB* (8.8%), *int-Tn* (80.7%) and *mefA* (0).

Conclusion: Majority of GBS contained PI-1+PI-2a. Hence presence of this pilus stabilizes the colonization, therefore designing a program for diagnosing and treatment of infected pregnant women seems to be necessary.

Keywords: *Streptococcus agalactiae*; Pili; Antibiogram; Women; Yazd

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INTRODUCTION

Streptococcus agalactiae, or group B Streptococcus (GBS) may colonize both genitourinary and gastrointestinal tract of human population (1-3). Different investigations revealed that 11-30% of pregnant women harbor this bacterium in their vagina

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and as a result their infant may get infected during the birth (2, 4). Investigation expressed that overall of 4% of infected infants develop sepsis, rarely pneumonia and meningitis before or after delivery (1, 5-7). The site for colonization of GBS is mucosal epithelial cells and for colonizing the bacterium needs an adherence factor (8). Three adherence factors have been recognized as capsule, surface protein and pili (9). Recently, ten serotypes of GBS have been recognized according to antigenic properties of their polysaccharide capsules as Ia, Ib, II ...VIII and IX (5, 10, 11). In addition to capsule, pili play an important role as adherence and pathogenicity factors for further successful colonization and invasion to the host tissue (1, 12). According to latest reports, three pilus island genes rendering the pilus-like structure on the surface of the GBS (PI-1, PI-2a, PI-2b). Pili are composed of three subunit; a back bone and two ancillary proteins (13, 14). More recently, investigations from European countries and USA expressed that GBS strain carry either PI-2a or PI-2b, but many have PI-1 (13, 15). Research performed in different societies represent that resistances of the GBS to selected antibiotics are noticeably increasing. This could be mainly due to uncontrolled of antibiotics utilization (16). The selected antibiotic for treatment of GBS is penicillin G (17) However, tetracycline, erythromycin and clindamycin are recommended for individuals who are allergic or infected with penicillin resistance GBS (18). Different investigation represent that some of GBS isolates are found resistance to one of the antibiotics ampicillin, vancomycin, penicillin and clindamycin (19, 20).

The aim of the present study was to determine the frequency of Pilus Island and antibiotic resistance profile of the perviously serotyped GBS isolated from pregnant women (4).

MATERIALS AND METHODS

Fifty seven GBS isolates which were previously collected (2015-2016), identified and serotyped were selected. These were subjected to pilus-island and antibiotic resistance determination using molecular technique. This study was considered and approved by Ethical Committee of Shahid Sadoughi University of Medical Sciences, Yazd, Iran [IR.SSU.MEDICINE.REC.1396.447]. All GBS isolates were re-inoculated on blood agar and incubated at 37°C for 24

hrs. We used purified DNA (GBS strain 389, 337) confirmed by Fanrong Kong from the centre for Microbiology Laboratory Services (New South Wales, Australia) as positive control and sterile water for negative control.

Genomic DNA extraction. The DNA was extracted using boiling method as described by Madzivhandila et al. (14). One ml of the microbial suspension was transferred into a sterile microtube and centrifuged at 3000 rpm for 10 min. The supernatant was removed and 1 ml of PBS was added to the pellet and centrifuged. This process was repeated three times. The pellet was then mixed with 100 µl of nuclease free water and incubated at 95°C in water bath. Following 10 min incubation, the supernatant was removed gently into another microtube and stored at -20°C for further molecular examination.

Detection of Pilus Island genes. The presence of GBS pilus island PI-1, PI-2a and PI-2b was detected by multiplex PCR as described by Martins et al. (15).

In order to confirm the PI-1 negative isolates that did not carry the pilus pathogenicity island or parts specific of its, using primers PI-1- All. All isolates that were negative for PI-1 gene were subjected to PCR with primers PI-1- All.

Antibacterial susceptibility testing. Sensitivity to penicillin (10 µg), tetracycline (30 µg), erythromycin (15 µg), clindamycin (2 µg), gentamycin (120 µg) and levofloxacin (5 µg) (MAST, England) was determined using Kirby Bauer test as described by CLSI (2018) (21). Briefly, McFarland 0.5 suspension of GBS was swabbed over the surface of Muller Hinton agar plate (Merck, Germany) contained 5% sheep blood. Then the disk containing each antibiotic was placed onto the inoculated surface. In addition inducible clindamycin resistance was detected using the D- Zone test according to CLSI 2018 guidelines (21). Following overnight incubation, the inhibition zone of each disk was measured and the isolate was interpreted as either susceptible, intermediate or resistance. Consistently *Streptococcus pneumoniae* (ATCC 49619) was used as a control sample.

Determination of resistance genes. All GBS strains were subjected to PCR technique for determination of resistance genes *ermB*, *mefA* as previously described (22). Also multiplex PCR was employed

for detection of *ermA*, *tetM*, *int-Tn* (22).

Statistical analysis. Chi-square test was assessed using SPSS version 20.

RESULTS

Amplification of pilus gene. As Table 1 and Fig. 1 shows, all isolates of GBS represent at least one pilus gene. Majority of the isolates (71.9%) contained the gene PI1+PI2a followed by PI2a (21.1%) and PI2b (7%). The gene PI1+ PI2b was not detected in any of the isolate. As Table 1 indicates, 33 (57.9%) of GBS were serotype III, among which, 30 (90.9%) were found to contain PI1+ PI2a and 3 isolates (9.1%) with PI2a gene.

Antimicrobial susceptibility. As Fig. 2 indicates, 51 (89.5%) of isolates were sensitive to penicillin, while 55 (96.5%) were resistant to tetracycline. Twelve isolates (21.1%) with positive D-zone were inducible resistance to clindamycin (iMLSB). Table 2 represents phenotypical and genotypical resistance to both macrolide and tetracycline. As it shows, 8 isolates were found to be phenotypically resistant to macrolide but 5 expressed *ermB* and 19 were *ermA* gene positive. Further consideration revealed that 11 (91.7%) out of 12 isolates contained *ermA* gene with serotype III and the remainder 1 isolate was found to be serotype Ia. Six isolates were resistance to both clindamycin and erythromycin (cMLSB). Among which, 5 isolates contained *ermB* and one *ermA* gene. Two (5%) of serotype III were resistance to erythromycin but sensitive to clindamycin (M phenotype) and 2 serotypes Ia and II were resistance to clindamycin but sensitive to erythromycin (L phenotype).

Correlation between the resistance gene of *tetM* (93%), *ermA* (33.3%), *ermB* (8.8%), *mefA* (0) and *int-Tn* (80.7%) with pili and different capsular serotype

are all shown in Table 3.

DISCUSSION

The present study was implicated one of the important virulence factor, Pilus- Island and tried to determine the correlation between the identified Pilus Island and the previous serotyped GBS in our area.



Fig. 1. Gel electrophoresis of multiplex PCR amplification products for determination of Pilus Island genes of GBS isolates. Lane 1 is DNA ladder (100 bp, Fermentas USA). PI1+PI2a (lane 3, 4, 6, 7 and 8). PI2a (Lane 2 and 5). PI2b is lane 9. Lane 10 is negative control.

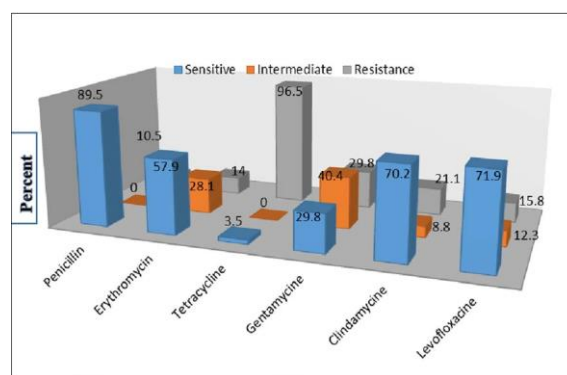


Fig. 2. Antibiotic susceptibility of Group B Streptococcus isolates

Table 1. Distribution of genes encoding Pilus- Island across serotypes

Pilus Island	Capsular Serotypes. n (%)					Total
	Ia	Ib	II	III	V	
PI1+PI2a	4 (50)	1 (100)	5 (35.7)	30 (90.9)	1 (100)	41 (71.9)
PI2a	1 (12.5)	0	8 (57.1)	3 (9.1)	0	12 (21.1)
PI2b	3 (37.5)	0	1 (7.1)	0	0	4 (7)
Total	8 (100)	1 (100)	14 (100)	33 (100)	1 (100)	57 (100)

P value= 0.000

In addition, all isolates were further retested to find out the frequency of the resistance to selected antibiotics. All GBS were found to contain one of two genes encoded pili's proteins (23). In addition, our study demonstrated that all isolates harbored at least one of PI gene alone or in combinations (Table 1).

As Table 1 shows, 41 (71.9%) of isolates contained PI-1+PI-2a. These results, however, concur with Margarit et al. (24) who reported that the most isolates they detected contained PI1+PI2a (46%). Although PII+PI2b were not observed in our investigation, prevalence of PI2a and PI2b were found to be 21.1% and 7% respectively. Different studies represented that majority of pili gene detected from GBS strain are PI2a (25, 26). They claimed that this gene may contribute to biofilm synthesis, but PII+PI2b does not play any function in biofilm synthesis. In a similar study, Rinaudo et al. showed that the most isolates of GBS (59.3%) harbor PII+PI2a genes. The results they obtained are not compromised with our finding in which PII+PI2a were found to be 71.9%. However, investigation concerning the role of PII+PI2a in GBS revealed that the presence of this pilus-island can enhance the virulence of species in both colonization and formation of biofilm (26).

Martins et al. in Spain examined 898 GBS strains

and reported that majority of isolates contained PII+PI2a with 49% but PI2b was the least (0.6%). Also they represented that 21% of the isolates were found to harbor PII+PI2b. Their results did not correspond with our finding and seems probably due to small number of cases we investigated (27).

Through our investigation, it was found that the majority of isolates contained PII+PI2a and belong to serotype III (30/57), followed in descending order by serotype II (5), Ia (4), Ib (1) and V (1). This figure of prevalence observed in our study, correspond with finding of Lue et al. who reported that 62/88 (70.5%) of PII+PI2a were serotype III (23). It is important to note that serotype III is more concerned with neonate meningitis (2).

Our finding also concur with those of Khodaei et al. who revealed that 67% of the GBS isolates were PII+PI2a and were mostly seen in serotype III (28).

Madzivhandila et al. in Africa demonstrated that the prevalence of pili type PII+PI2b was as 45.1% followed by PI2a (29.8%), PI1+PI2a (24.8%) and PI2b with 0.2%. In addition, they expressed that the presence of PII+PI2b can cause invasive disease and PII+PI2a contribute the GBS for colonization. They showed that PII+ PI2b were seen with serotype III and PI2a were found mostly in serotype Ia (29).

Margarit et al. found the most common serotypes of Ib, II and V contained two separate pili, whereas, majority of pilus- island PI2a was found in serotype Ia (24).

Fig. 2. indicates the result of susceptibility of detected GBS against selected antibiotics. Although the majority of isolates were resistance to tetracycline (96.5%), they showed the lowest resistance to penicillin (10.5%). Majority of the isolates harbored gene *tetM* (93%) followed by *int-tn* (80.7), *mefA* (0), *ermA* (33.3%) and *ermB* (8.8%). In a review study, Gizachew (30) reported 36.6% resistance rate for penicillin among 1974 positive GBS detected from pregnant women. In a another study from Iran, 88.5% of iso-

Table 2. Resistance phenotype and genotype (Macrolide – Tetracycline, n=57)

Macrolide- resistance phenotype (n)	8
Macrolide- resistance genotype (n)	
<i>ermB</i>	5
<i>ermA mefA</i>	19
<i>ermA+ermB</i>	0
Tetracycline resistance phenotype (n)	0
Tetracycline- resistance genotype (n)	55
<i>tetM</i>	
<i>Int-Tn</i>	53
	46

Table 3. Distribution of serotypes and Pilus- Island among antimicrobial resistance genes of GBS isolates (N=57)

Antibiotic resistance genotype (n/%)	Serotypes (n/%)	Pilus island (n/%)
<i>ermA</i> (19/ 33.3)	Ia (1/5.3), III (18/94.7)	PI2a (1/5.3), PI-1+PI2a (18/94.7)
<i>ermB</i> (5/ 8.8)	Ia (1/20), Ib (1/20), II (1/20), III (1/20), V (1/20)	PI2a (2/40), PII+PI2a (3/60)
<i>tetM</i> (53/ 93)	Ia (6/11.3), Ib (1/1.9), II (13/24.5), III (32/60.4), V (1/1.9)	PI2a (10/18.9), PI2b (3/5.7), PII+PI2a (40/75.7)
<i>int-Tn</i> (46/ 80.7)	Ia (6/13), II (8/17.4), III (31/67.4), V (1/2.2)	PI2a (6/13), PI2b (3/6.5), PII+PI2a (37/80.4)
<i>mefA</i> (0)	0	0

lates GBS were found to be sensitive to penicillin (33).

As Table 2 indicates, 8 GBS isolates were found resistance to erythromycin but, however, 19 isolates were positive for *ermA*. This controversy may be explained that this gene is not expressed yet (32).

When our results were compared to Martins et al. it was found that 85.8% of their obtained isolates were resistance to tetracycline. They also reported that only 16.1% and 14.2% of the GBS were resistance to erythromycin and clindamycin respectively. Concerning the gene distribution, we found that 93% of the isolates contain *tetM* gene, whereas they reported that 97.3% of their detected GBS have had the above gene (33).

Another study conducted by Heelan et al. revealed that 85% of the isolates were tetracycline resistance which is almost similar with our finding. They also expressed that 22% of the isolates were resistant to erythromycin and 100% of isolates contained erythromycin resistance gene. Important point was that they showed cMLSB isolates carried the gene *ermA* and *ermB* which was similar to our result. Similarly, they found that the phenotype of iMLSB was directly related to gene *ermA* activity (34).

tetM is recognized as a most prevalent gene in tetracycline resistant Gram positive bacteria and is frequently associated with conjugate elements which belong to the family of Tn916. Today, it is well known that *int-Tn* encodes the integrase of Tn916 continuously and it is found in 88% of strains with *tetM* (22). Our investigation revealed that 80.7% of isolated GBS contained *int-Tn*. This finding indicates that tetracycline resistance GBS is probably due to their achievement to transposon of Tn916. As Table 3 shows, there was not any *mefA* gene among our tested GBS. It seems that resistance of isolates to erythromycin is accomplished by transposon (*ermB*, *ermA*) and or through efflux pump like *mefA* gene.

CONCLUSION

Results obtained from this investigation allow us to conclude that majority of isolated GBS contain pili PII+PI2a and resistance of these isolates to clindamycin, erythromycin and tetracycline are constantly increasing. Since presence of these pili is believed to enhance the bacterial colonization on host's target tissue, diagnostic program followed by treatment of infected pregnant women during pregnancy is

strongly recommended.

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REFERENCES

1. Springman AC, Lacher DW, Waymire EA, Wengert SL, Singh P, Zadoks RN, et al. Pilus distribution among lineages of group B streptococcus: an evolutionary and clinical perspective. *BMC Microbiol* 2014;14:159.
2. Sadeh M, Firouzi R, Derakhshandeh A, Khalili MB, Kong F, Kudinha T. Molecular characterization of *Streptococcus agalactiae* isolates from pregnant and non-pregnant women at Yazd University Hospital, Iran. *Jundishapur J Microbiol* 2016;9(2): e30412.
3. Manshadi SAD, Alizadeh R, Salehi M, Seifi A, Seifi M. Bilateral septic arthritis of the knee caused by group B streptococci: a case report. *Iran J Microbiol* 2019;11:187-190.
4. Najarian N, Khalili MB, Astani A, Vakili M, Sadeh M. Serotype determination of *Streptococcus agalactiae* detected from vagina and urine of pregnant women in Yazd, Iran-2015. *IJML* 2018;5:49-57.
5. Nazari A, Khalili MB, Astani A, Vakili M, Sadeh M, Mojibiyani M, et al. Determination of genotypes of *Streptococcus agalactiae* isolated from both urine and vagina of pregnant women referred to gynecology clinics of Yazd, Iran-2015. *IJML* 2017;4:180-188.
6. Dehbashi S, Pourmand MR, Mashhadi R. Characterization of Afb, a novel bifunctional protein in *Streptococcus agalactiae*. *Iran J Microbiol* 2016;8:73-79.
7. Heath PT, Balfour GF, Tighe H, Verlander NQ, Lamagni TL, Efstratiou A, et al. Group B streptococcal disease in infants: a case control study. *Arch Dis Child* 2009;94:674-680.
8. Tamura G, Kuypers J, Smith S, Raff H, Rubens C. Adherence of group B streptococci to cultured epithelial cells: roles of environmental factors and bacterial surface components. *Infect Immun* 1994;62:2450-2458.
9. Wessels MR, Rubens CE, Benedi VJ, Kasper DL. Definition of a bacterial virulence factor: sialylation of the group B streptococcal capsule. *Proc Natl Acad Sci U S A* 1989;86:8983-8987.
10. Imperi M, Pataracchia M, Alfarone G, Baldassarri L, Orefici G, Creti R. A multiplex PCR assay for the direct

- identification of the capsular type (Ia to IX) of *Streptococcus agalactiae*. *J Microbiol Methods* 2010;80:212-214.
11. Hanh TQ, Van Du V, Hien PT, Chinh DD, Loi CB, Dung NM, et al. Prevalence and capsular type distribution of group B Streptococcus isolated from vagina of pregnant women in Nghe An province, Vietnam. *Iran J Microbiol* 2020;12:11-17.
 12. Vengadesan K, Ma X, Dwivedi P, Ton-That H, Narayana SV. A model for group B Streptococcus pilus type 1: the structure of a 35-kDa C-terminal fragment of the major pilin GBS80. *J Mol Biol* 2011;407:731-743.
 13. Sharma P, Lata H, Arya DK, Kashyap AK, Kumar H, Dua M, et al. Role of pilus proteins in adherence and invasion of *Streptococcus agalactiae* to the lung and cervical epithelial cells. *J Biol Chem* 2013;288:4023-4034.
 14. Madzivhandila M, Adrian PV, Cutland CL, Kuwanda L, Madhi SA, Team PT. Distribution of pilus islands of group B streptococcus associated with maternal colonization and invasive disease in South Africa. *J Med Microbiol* 2013;62:249-253.
 15. Martins ER, Melo-Cristino J, Ramirez M. Evidence for rare capsular switching in *Streptococcus agalactiae*. *J Bacteriol* 2010;192:1361-1369.
 16. Phares C, Lynfield R, Farley M, Mohle-Boetani J, Harrison L, Petit S, et al. Epidemiology of invasive group B streptococcal disease in the United States, 1999-2005. *JAMA* 2008;299:2056-2065.
 17. Wang YH, Su LH, Hou JN, Yang TH, Lin TY, Chu C, et al. Group B streptococcal disease in nonpregnant patients: emergence of highly resistant strains of serotype Ib in Taiwan in 2006 to 2008. *J Clin Microbiol* 2010;48:2571-2574.
 18. Gherardi G, Imperi M, Baldassarri L, Pataracchia M, Alfarone G, Recchia S, et al. Molecular epidemiology and distribution of serotypes, surface proteins, and antibiotic resistance among Group B streptococci in Italy. *J Clin Microbiol* 2007;45:2909-2916.
 19. Sadowy E, Matynia B, Hryniewicz W. Population structure, virulence factors and resistance determinants of invasive, non-invasive and colonizing *Streptococcus agalactiae* in Poland. *J Antimicrob Chemother* 2010;65:1907-1914.
 20. Daramroodi AK, Keshavarzi F, Raissi F. The investigation of antibiotic resistance and rapid detection of group B Streptococcus (Bca) from vaginal specimens of pregnant women by colony PCR method. *J Bas Res Med Sci* 2018; 5:27-32.
 21. Clinical and Laboratory Standards Institute (CLSI). Performance Standards for Antimicrobial Susceptibility Testing: Approved Twenty-: Document M100-S28. Wayne, PA, USA: CLSI 2018.
 22. Poyart C, Jardy L, Quesne G, Berche P, Trieu-Cuot P. Genetic basis of antibiotic resistance in *Streptococcus agalactiae* strains isolated in a French hospital. *Antimicrob Agents Chemother* 2003;47:794-797.
 23. Lu B, Chen X, Wang J, Wang D, Zeng J, Li Y, et al. Molecular characteristics and antimicrobial resistance in invasive and noninvasive Group B Streptococcus between 2008 and 2015 in China. *Diagn Microbiol Infect Dis* 2016;86:351-357.
 24. Margarit I, Rinaudo CD, Galeotti CL, Maione D, Ghezzi C, Buttazzoni E, et al. Preventing bacterial infections with pilus-based vaccines: the group B streptococcus paradigm. *J Infect Dis* 2009;199:108-115.
 25. Xia FD, Mallet A, Caliot E, Gao C, Trieu-Cuot P, Dramsi S. Capsular polysaccharide of Group B Streptococcus mediates biofilm formation in the presence of human plasma. *Microbes Infect* 2015;17:71-76.
 26. DeLeo FR, Rinaudo CD, Rosini R, Galeotti CL, Berti F, Necchi F, et al. Specific involvement of Pilus type 2a in biofilm formation in Group B Streptococcus. *PLoS One* 2010;5(2): e9216.
 27. Martins E, Andreu A, Melo-Cristino J, Ramirez M. Distribution of Pilus Islands in *Streptococcus agalactiae* that cause human infections: insights into evolution and implication for vaccine development. *Clin Vaccine Immunol* 2013;20:313-316.
 28. Khodaei F, Najafi M, Hasani A, Kalantar E, Sharifi E, Amini A, et al. Pilus-encoding islets in *S. agalactiae* and its association with antibacterial resistance and serotype distribution. *Microb Pathog* 2018;116:189-194.
 29. Madzivhandila M, Adrian PV, Cutland CL, Kuwanda L, Madhi SA, The Po PSTT. Distribution of pilus islands of group B streptococcus associated with maternal colonization and invasive disease in South Africa. *J Med Microbiol* 2013;62:249-253.
 30. Gizachew M, Tiruneh M, Moges F, Tessema B. *Streptococcus agalactiae* maternal colonization, antibiotic resistance and serotype profiles in Africa: a meta-analysis. *Ann Clin Microbiol Antimicrob* 2019;18: 14.
 31. Ghanbarzadeh N, Mehramiz M, Gannadkafi M, Namaei MH. The prevalence of group B streptococcus rectovaginal colonization and antimicrobial susceptibility pattern among pregnant women: a descriptive-analytical study. *Mod Care J* 2017;14(3):e66391.
 32. Bolukaoto JY, Monyama CM, Chukwu MO, Lekala SM, Nchabeleng M, Maloba MR, et al. Antibiotic resistance of *Streptococcus agalactiae* isolated from pregnant women in Garankuwa, South Africa. *BMC Res Notes* 2015;8: 364.
 33. Martins ER, Pedroso-Roussado C, Melo-Cristino J, Ramirez M, The Portuguese Group for the Study of Streptococcal Infections. *Streptococcus agalactiae* causing neonatal infections in portugal (2005-2015): diversification and emergence of a CC17/PI-2b multidrug resistant sublineage. *Front Microbiol* 2017;8:499.
 34. Heelan JS, Hasenbein ME, McAdam AJ. Resistance of group B Streptococcus to selected antibiotics, including erythromycin and clindamycin. *J Clin Microbiol* 2004;42:1263-1264.