



The frequency of Epstein-Barr virus among hemodialysis patients, Ahvaz, Iran

Rahil Nahid Samiei¹, Shahab Mahmoudvand¹, Somayeh Shokri¹, Manoochehr Makvandi^{1,2*}, Heshmatollah Shahbazian³, Roya Pirmoradi¹, Shokouh Shayanpur⁴, Kimia Makvandi⁵, Sepideh Nowrozi⁵

¹Department of Virology, School of Medicine, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran ²Infectious and Tropical Diseases Research Center, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran

³Department of Nephrology, Golestan Hospital, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran

⁴Nephrology Department, Imam Khomeini Hospital, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran

⁵School of Medicine, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran

Received: March 2018, Accepted: December 2018

ABSTRACT

Background and Objectives: Epstein-Barr virus (EBV) has infected more than 90% of adults worldwide. EBV infection is asymptomatic in healthy individuals and is controlled by a robust immune response while in individuals with weakened immunesystems including Hemodialysis (HD) patients and transplant recipients leads to serious illnesses. This study was aimed to investigate the frequency of EBV among the HD patients.

Materials and Methods: The cross-sectional study was carried out on 84 HD patients. These sera were checked for anti-EBV (VCA) IgG Ab assessment using enzyme-linked immunosorbent assay (ELISA). The DNA was extracted from the sera samples and tested for EBV DNA using nested PCR.

Results: 52/84 (61.9%) of HD were males and 32/84 (38.1%) were females. The average age of participants was varying from 18 to 85 years while the mean age was 52 ± 1.57 SD years. 81 of 84 (96.42%); including 49/52 (94.23%) male and 32/32 (100%) female, were positive for anti-EBV (VCA) IgG antibody while 3 (3.58%) were negative. No significant differences were observed between the subjects regarding gender (P=0.28). EBV DNA was detected in 7 (8.33%) individuals, including 6 (11.53%) and 1 (3.12%) in male and female, respectively (P=0.24).

Conclusion: Our study results showed that high prevalence of anti-EBV (VCA) IgG antibody (96.42%) were observed among the HD patients. Although the status of EBV latency was not performed, but it seems many of these patients are at risk of EBV-reactivation during the organ transplantation. As a result, it is recommended that the detection of EBNA-1 gene as a marker of EBV latency should be implemented for all HD patients to prevent EBV reactivation during organ transplantation.

Keywords: Epstein-Barr virus; Hemodialysis patients; Enzyme-linked immunosorbent assay

*Corresponding author: Prof. Manoochehr Makvandi, Department of Virology, School of Medicine, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran; Infectious and Tropical Diseases Research Center, Ahvaz

Jundishapur University of Medical Sciences, Ahvaz, Iran.

Tel: +98-9166181683 Fax: +98-6133738313

Email: manoochehrmakvandi29@yahoo.com

INTRODUCTION

Epstein-Barr virus (EBV), a member of the herpesvirus family, is one of the most common human infections that infects more than 95% of the world populations (1). It has been implicated in several diseases, including infectious mononucleosis, African Burkitt lymphomas (BL), Hodgkin's lymphoma, B-cell lymphomas of immunocompromised, nasopharyngeal carcinomas (NPC), stomach cancer. Most individuals contract EBV infection in early adulthood (2). The transmission of EBV occurs mainly via contact with saliva. However, the virus can be transmitted through sexual contact, blood transfusions, and organ transplantations (3, 4). Hemodialysis (HD) is a process of purifying the blood of a person whose kidneys are not working normally, especially patients in end-stage renal failure. Renal transplantation is considered the treatment of choice for these patients (5). Patients in end-stage renal failure have severe alterations in cell mediated immunity that increase their risk of contracting opportunistic viral infections such as EBV infection (6). EBV is most likely to cause problems when the immune system is suppressed by disease (7). Therefore, immunocompromised hemodialysis patients and renal transplant patients are at high risk for EBV infection. Like all herpesviruses, after primary infection, EBV establishes a persistent infection in almost all infected host, which may be responsible for the development of several diseases. The virus can cause a latent infection within B lymphocytes, allowing the virus to evade the host immune response (8). Latent EBV in B cells can be reactivated to switch to lytic replication, especially in individuals with weakened immune systems (9, 10). Post-transplant lymphoproliferative (PTLD) is one of the most important EBV-associated complication. PTLD is a severe complication of solid organ that occur after a transplant, a leading life-threatening malignancy in the transplant population. It can develop in people who are taking immunosuppressive drugs to prevent rejection of an organ. EBV is the main driver of PTLD, particularly in patients with impaired immunosurveillance against EBV, and is contributes to the pathogenesis of PTLD in more than 70% of cases (11, 12). Therefore, the PTLD occurrence is preceded by increased number of latently infected B-cells and EBV reactivation (8). The virus-associated various tumors mainly encompass latently infected B-lymphocytes in which EBV-encoded growth-transformation-associated proteins are expressed (13). Therefore, to prevention of EBV complications, hemodialysis patients should be routinely tested for EBV Viral Capsid Antigen (VCA) IgG antibody and EBV DNA before transplantation. The aim of this study was to evaluate serological and molecular status of EBV infection in the HD patients.

MATERIALS AND METHODS

Samples collection. In this cross-sectional study, the sera of 84 (including 32 females and 52 males) HD patients who referred to Golestan hospital were collected during October 2014 to November 2014. Their sera were stored at -20 before use.

EBVspecific IgG antibody detection by Enzyme-linked Immunosorbent Assay (ELISA). Sera were tested for the presence of EBV Viral Capsid Antigen (VCA) IgG antibody by ELISA kit (DIAPRO, diagnostic, Milan, Italy) according to the manufacturer's instruction. Cut-off was defined with positive and negative control sera that were included in each assay, according to the manufacturer's instruction.

DNA extraction. DNA was extracted from serum using High Pure PCR Template Preparation kit (Roche, Germany) according to the manufacturer's instructions. The extracted DNA was reanalyzed for the presence of the EBVDNA using nested PCR.

EBV DNA detection by nested PCR. All samples were subjected to nested PCR for detection of EB-VDNA, using specific primers, EBV outer Forward: 5'-TGGAAAC CCGTCA CTCTC-3', Reverse: 5'-TA-ATGGCATAGGTGGAATG-3' primers were used for first run and EBV Inner Forward: 5'-AGGGAT-GCCTGGACACAAGA-3', Reverse: 5'-GCCTCG-GTTGTGACAGAG-3' primers for second run (14). Nested PCR was performed in a total volume of 25 μL, containing the 5 μl extracted DNA, 0.2 μl MgCl, 25 mM,0.5 µl deoxyribonucleotide triphosphates solution 10 µM (Roche, Germany), 2.5 µl PCR buffer 10× (Roche, Germany), 1U Taq DNA polymerase (Roche, Germany), 1 µl from each primer, and 14.5 µl distilled water. Following thermal conditions was carried out: For the first round of PCR program: initial incubation at 94°C for 5 min followed by 35 cycles including 95°C for 45 sec, annealing at 50°C for 45 sec and extension at 72°C for 45 sec, and final extension at 72°C for 10 min. The second round PCR was the same as the first round PCR but annealing temperature was 53°C.

Then PCR product was subjected to 2% agarose gel stained with SYBR Safe DNA gel stain. The bands were visualized using UV transilluminator.

Statistical analysis. The collected data were analyzed with SPSS 16 package program (SPSS Inc., Chicago, IL, USA) and Fisher's exact test was used to assess the rate of EBV antibody and EBVDNA among the gender detection calculate. A P-value below 0.05 was considered statistically significant.

RESULTS

Of 84 HD patients 52 (61.9%) were male and 32 (38.1%) were female. The average age of participants was varying from 18 to 85 years with mean ages of 52 ± 1.57 SD years. 81 (96.42%) subjects were shown positive for anti-EBV IgG antibody while 3 (3.58%) were negative. Overall the prevalence of anti-EBV IgG was 96.42%. Among them 49/52 (94.23%) male and 32/32 (100%) female were positive for anti-EBV IgG antibody. No significant differences were observed between the subjects regarding gender (P=0.28). EBV DNA was detected in 7/84 (8.33%) (Table 1).

DISCUSSION

Immunosuppressed patients may flunk to create an effective immune response against EBV. This

Table 1. EBV DNA distribution in male and female HD patients

Gender	Positive	Negative	P-value
Male	6	46	
(n=52)	(11.53%)	(88.47%)	
Female	1	31	P=0.24
(n=32)	(3.12%)	(96.88%)	
Total	7	77	
(n=84)	(8.33%)	(91.67%)	

may result in to a persistent infection, which may be account for the creation of PTLD, Burkitt lymphoma, and Large B cell lymphoma, are well-known life-threatening complication of solid organ transplantation (15, 16).

In the current study, the rate of positivity for anti-EBV (VCA) IgG antibody was 81/84 (96.42%). Also, EBV DNA was detected in 7/84 (8.33%). The rate of EBV DNA among the male (11.53%) and female (3.12%) was not significant (P=0.24). No significant differences was observed in EBV IgG antibody in male (94.23%) and female (100%) individuals (P=0.28).

In study conducted by Verghese et al in USA, EBV DNA was detected in 32 /95 (34%) pre-transplant (17). Study performed by Nikoobakht et al. demonstrated that the frequency of positive EBV DNA in pre-transplant saliva samples was 44.1% increasing to 67.6%, after transplantation (18). This controversy in EBV DNA positive cases may come from the differences in genetic background of the patients and differences in EBV distribution in different country. The prevalence of anti-EBV IgG in hemodialysis patients in Croatia was 98% (19). In another study performed in Cyprus, the prevalence of EBV IgG antibodies among hemodialysis patients was 94% (20). Saghafi et al. reported that the prevalence of EBV IgG antibody is 100% among adult potential donors and recipients (21). Our results were consistent with recent studies. In study conducted by Beladi Mousavi, EBV IgG antibody was positive in 70% of recipients and 52% of donors but there was no statistically significant difference between males and females (p=0.94) (22). The present study found no significant difference in EBV IgG antibody seroprevalence between genders, which is similar to the result of study conducted by Beladi Mousavi among hemodialysis patients. In study performed in Iran, 15.5% of the renal transplant recipients were positive for EBV infection (23).

It is noteworthy that EBV has been identified as a cofactor in the pathogenesis of a significant proportion of PTLD (11). Furthermore, the virus may lead to graft rejection (24, 25). It has also been reported some of EBV associated lymphomas after solid organ transplant (26). Currently, there is no licensed vaccine to prevent EBV infection. Several clinical trials for a vaccine were conducted in the world (27). Therefore, it seems that screening is required to reduce the further complications of EBV infection in

hemodialysis patients. Detection of EBNA-1 protein or EBNA-1 gene is marker of the EBV latent infection. There are serologic tests for assessing the status of EBV prior infection and reactivation. Antibodies to EBNA-1, Epstein-Barr virus nuclear antigen 1, can be measured to help diagnose prior infection while early antigen-diffuse (EA-D) IgG, can act as a marker of EBV reactivation (28). As well as, EBV viral load test, can help as a predictor of EBV-related post-transplant lymphoproliferative disorders in transplant patients (29). Treatment with immunosuppressive drugs may lead to EBV reactivation. Thus, to prevent reactivation before starting immunosuppression therapeutic strategies such as valganciclovir should be used in immunocompromized patient populations who are at high risk of EBV reactivation (30).

In sum, the results of the present study confirm a high prevalence of EBV IgG seropositivity among HD patients. As a result, due to the clinical importance of EBV reactivation and its serious complications in immunosuppressed patients, it is suggested that further studies with more sample size to be conducted to determine the role of EBV in hemodialysis patients. Although the status of EBV latency was not evaluated in the present study but it requires further investigation. Thus, it is recommended that the detection of EBNA-1 antibody, EBV viral load and EA-D IgG as a marker of EBV latency and EBV reactivation should be implemented for all HD patients prior to kidney transplant to prevent EBV reactivation and EBV complication consequences.

ACKNOWLEDGEMENTS

This study was financially supported by Ahvaz Jundishapur University of Medical Sciences (Grant no: 92-105).

REFERENCES

- Chabay P, Preciado MV. Epidemiology of Epstein-Barr virus-associated pediatric lymphomas from Argentina. Bol Med Hosp Infant Mex 2016;73:47-54.
- 2. Maeda E, Akahane M, Kiryu S, Kato N, Yoshikawa T, Hayashi N, et al. Spectrum of Epstein-Barr virus-related diseases: a pictorial review. *Jpn J Radiol* 2009;27:4-19.

- Pagano JS. Is Epstein-Barr virus transmitted sexually? JID 2007;195:469-470.
- Sitki-Green D, Covington M, Raab-Traub N. Compartmentalization and transmission of multiple epstein-barr virus strains in asymptomatic carriers. *J Virol* 2003;77:1840-1847.
- Abecassis M, Bartlett ST, Collins AJ, Davis CL, Delmonico FL, Friedewald JJ, et al. Kidney transplantation as primary therapy for end-stage renal disease: a national kidney foundation/ kidney disease outcomes quality initiative (NKF/KDOQITM) conference. *Clin J Am Soc Nephrol* 2008;3:471-480.
- Kato S, Chmielewski M, Honda H, Pecoits-Filho R, Matsuo S, Yuzawa Y, et al. Aspects of immune dysfunction in end-stage renal disease. *Clin J Am Soc Nephrol* 2008;3:1526-1533.
- Merlo A, Turrini R, Dolcetti R, Martorelli D, Muraro E, Comoli P, et al. The interplay between Epstein-Barr virus and the immune system: a rationale for adoptive cell therapy of EBV-related disorders. *Haematologica* 2010;95:1769-1777.
- Habib M, Buisson M, Lupo J, Agbalika F, Socié G, Germi R, et al. Lytic EBV infection investigated by detection of Soluble Epstein-Barr virus ZEBRA in the serum of patients with PTLD. Sci Rep 2017; 7: 10479.
- Michallet M, Sobh M, Ranchon F, Leroy S, Barraco F, Thomas X, et al. Epstein-Barr Virus (EBV) reactivation, its treatment with Rituximab and their impact on relapse after allogeneic hematopoietic stem cell transplantation for hematological malignancies. *Blood* 2016;128:3695.
- Wu C-C, Fang C-Y, Cheng Y-J, Hsu H-Y, Chou S-P, Huang S-Y, et al. Inhibition of Epstein-Barr virus reactivation by the flavonoid apigenin. *J Biomed Sci* 2017; 24: 2.
- San-Juan R, Comoli P, Caillard S, Moulin B, Hirsch HH, Meylan P. Epstein-Barr virus-related post-transplant lymphoproliferative disorder in solid organ transplant recipients. *Clin Microbiol Infect* 2014;20:109-118.
- 12. Petrara MR, Giunco S, Serraino D, Dolcetti R, De Rossi A. Post-transplant lymphoproliferative disorders: from epidemiology to pathogenesis-driven treatment. *Cancer Lett* 2015;369:37-44.
- Yajima M, Kanda T, Takada K. Critical role of Epstein-Barr virus (EBV)-encoded RNA in efficient EBV-induced B-Lymphocyte growth transformation. *J Virol* 2005;79:4298-4307.
- 14. Hassan R, White LR, Stefanoff CG, de Oliveira DE, Felisbino FE, Klumb CE, et al. Epstein-Barr virus (EBV) detection and typing by PCR: a contribution to diagnostic screening of EBV-positive Burkitt's lymphoma. *Diagn Pathol* 2006;1:17.
- 15. Green M, Michaels MG. Epstein-Barr virus infection and posttransplant lymphoproliferative disorder. *Am J*

- Transplant 2013;13:41-54.
- 16. Elisa C, Isabella G, Federica M, Germana L, Cristina R, Alessandra L, et al. Viral load of EBV DNAemia is a predictor of EBV-related post-transplant lymphoproliferative disorders in pediatric renal transplant recipients. *Pediatr Nephrol* 2017; 32:1433-1442.
- Verghese PS, Schmeling DO, Filtz EA, Grimm JM, Matas AJ, Balfour HH. Transplantation of solid organ recipients shedding Epstein-Barr virus DNA pre-transplant: A prospective study. *Clin Transplant* 2017;31(11):e13116.
- Nikoobakht M, Beitollahi J, Nikoobakht N, Aloosh M, Sahebjamee M, Rezaeidanesh M, et al. Evaluation of Epstein–Barr virus load in Saliva before and after renal transplantation. *Transplant Proc* 2011;43:540-542.
- Vilibić-Čavlek T, Kolarić B, Bogdanić M, Tabain I, Beader N. Herpes group viruses: a seroprevalence study in hemodialysis patients. *Acta Clin Croat* 2017; 56:255-261.
- Elie D, Dana K, Efthychia G, Astero C, Anastasia L, Marios P, et al. Evaluation of Epstein-Barr virus-specific antibodies in Cypriot multiple sclerosis patients. *Mol Immunol* 2019;105:270-275.
- 21. Saghafi H, Qorashi M, Heidari A. Is screening for IgG antibody to cytomegalovirus and Epstein-Barr virus infections mandatory in potential renal transplant recipients and donors in Iran? *Transplant Proc* 2009;41:2761-2763.
- 22. Beladi Mousavi SS. Do we need to screen our patients for EBV IgG antibody before kidney transplantation? *Nephrourol Mon* 2011;3:122-124.
- 23. Hasannia T, Moosavi Movahed SM, Vakili R, Rafatpanah H, Hekmat R, Valizadeh N, et al. Active CMV and EBV infections in renal transplant recipients with unexplained fever and elevated serum creatinine. *Ren Fail* 2016;38:1418-1424.

- Babel N, Schwarzmann F, Prang N, Jaeger M, Wolf H, Kern F, et al. Association between Epstein-Barr virus infection and late acute transplant rejection in longterm transplant patients. *Transplantation* 2001;72:736-739
- 25. Ahya VN, Douglas LP, Andreadis C, Arnoldi S, Svoboda J, Kotloff RM, et al. Association between elevated whole blood Epstein–Barr virus (EBV)-encoded RNA EBV polymerase chain reaction and reduced incidence of acute lung allograft rejection. *J Heart Lung Transplant* 2007;26:839-844.
- Rohr JC, Wagner HJ, Lauten M, Wacker HH, Juttner E, Hanke C, et al. Differentiation of EBV-induced post-transplant Hodgkin lymphoma from Hodgkin-like post-transplant lymphoproliferative disease. *Pediatr Transplant* 2008;12:426-431.
- 27. Sokal EM, Hoppenbrouwers K, Vandermeulen C, Moutschen M, Léonard P, Moreels A, et al. Recombinant gp350 vaccine for infectious mononucleosis: a phase 2, randomized, double-blind, placebo-controlled trial to evaluate the safety, immunogenicity, and efficacy of an Epstein-Barr virus vaccine in healthy young adults. *J Infect Dis* 2007;196:1749-1753.
- Martinez OM, Krams SM. The immune response to Epstein Barr virus and implications for posttransplant lymphoproliferative disorder. *Transplantation* 2017;101:2009-2016.
- Colombini E, Guzzo I, Morolli F, Longo G, Russo C, Lombardi A, et al. Viral load of EBV DNAemia is a predictor of EBV-related post-transplant lymphoproliferative disorders in pediatric renal transplant recipients. *Pediatr Nephrol* 2017;32:1433-1442.
- 30. Gill H, Hwang Y-Y, Chan TSY, Pang AWK, Leung AYH, Tse E, et al. Valganciclovir suppressed Epstein Barr virus reactivation during immunosuppression with alemtuzumab. *J Clin Virol* 2014;59:255-258.