



Detection of methicillin-resistant *Staphylococcus aureus* (MRSA) in clinical samples of patients with external ocular infection

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ABSTRACT

Background and Objectives: *Staphylococcus aureus* is the main Gram-positive bacteria isolated from patients with ocular infections. Herein, we describe the pattern of antibiotic resistance, presence of resistance genes including *ermA*, *ermB*, *ermC*, *msrA*, *mecA* and the *pvl* cytotoxin gene in *S. aureus* isolates collected from patients with external ocular infection.

Materials and Methods: In this study, 8 *S. aureus* isolates were collected from 81 patients that suffered from eye damage. Antibacterial susceptibility of isolates was determined using the Kirby-Bauer disk diffusion method. Resistance genes including *ermA*, *ermB*, *ermC*, *msrA*, *mecA* and the *pvl* virulence gene were detected by PCR method. Staphylococcal cassette chromosome *mec* (SCC*mec*) in MRSA isolates were detected by the multiplex-PCR method.

Results: Three isolates were resistant to cefoxitin which is considered MRSA. The *mecA* gene was identified in MRSA isolates. SCC*mec* type IV and the *pvl* gene were detected in one of the MRSA isolates that was recovered from a diabetic patient. **Conclusion:** The emergence of *S. aureus* isolates belonging to SCC*mec* type IV and *pvl* gene among patients with ocular infection is very serious; therefore, identify genetic characterization of MRSA isolates for empirical therapy and infection control is very important.

Keywords: Eye infection, Methicillin resistance in Staphylococcus aureus, Panton-Valentine leukocidin

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INTRODUCTION

Staphylococcus aureus causes a wide range of infections including bacteremia as well as pleuropulmonary, skin and soft tissue infections (1, 2). This bacterium is also responsible for many types of ocular infections including keratitis, conjunctivitis, endophthalmitis and blepharitis (2). Some virulence factors such as Panton-valentine leukocidin (PVL) as well as antibiotic resistance have an important role in increasing tissue damage and failure in antibiotic therapy, respectively (1-3). There are many reports about ocular infection due to methicillin-resistant S. aureus (MRSA) isolates (2). In the past decade, infections by MRSA strains have increased in hospital settings and communities (2). MRSA isolates are commonly resistant to a wide range of antibiotics such as aminoglycosides, erythromycin, tetracycline and fluoroquinolones (1-3). The mecA gene is on the staphylococcal cassette chromosome mec (SCCmec) genetic element that causes resistance to β-lactam antibiotics (3-5). SCCmec type IV usually is related to community-associated-MRSA (CA-MRSA) and other SCCmec types including SCCmec types I, II and III related to hospital-acquired MRSA (HA-MRSA) isolates (5, 6). The PVL is a cytotoxin virulence factor produced by some S. aureus commonly related to CA-MRSA isolates and causes cell destruction and tissue necrosis (6). Antibiotics such as erythromycin and tetracycline were used for treatment of ocular infections by S. aureus (1). Activity of efflux pumps such as msr (A/B) and methylation of the ribosomal drug binding site by methylase enzymes encoded by erm (A, B, C) genes are important resistance mechanisms to macrolide antibiotics (3, 7). Herein, we described antibiotic susceptibility pattern, persence of msrA/B, ermA, ermB, ermC, mecA, SCCmec type and the pvl genes among S. aureus isolates recovered from patients with external ocular infection in Tehran, Iran.

MATERIALS AND METHODS

Bacterial isolates. In a cross-sectional study, from April 2016 to September 2016, 8 *S. aureus* isolates were collected from clinical samples of 81 different patients suffering from eye damage and purulent discharge admitted to Shahid Labafi Nejad Hospital in Tehran, Iran. Bacterial isolates were considered as *S. aureus* using standard microbiological tests in-

cluding Gram staining, catalase reaction, coagulase production, β-hemolysis on blood agar base enriched with 5% sheep blood, DNase production and mannitol fermentation on mannitol salt agar media (6). All culture media used in this investigation were supplied by HiMedia, Co, India. Finally, isolates confirmed by amplification of the *nuc* gene by the PCR technique as described previously (6).

Antibacterial susceptibility tests. Antibacterial susceptibility of isolates to amikacin (30 μg), ciprofloxacin (5 μg), clindamycin (2 μg), erythromycin (15 μg), gentamicin (10 μg), linezolid (30 μg), tetracycline (30 μg) and trimethoprim/sulfamethoxazole (1.25/23.75 μg) were determined using Kirby-Bauer disk diffusion method on Mueller hinton agar media (CONDA, Co, Spain) (8). Brain hearth infusion agar (BHI; Difco, USA) containing 6 μg/mL vancomycin was used for detection of vancomycin resistant isolates (8). MRSA and inducible clindamycin resistance phenotypes were determined using the CLSI recommendations (8). The *S. aureus* ATCC 29213 was used as the control strain in antibacterial susceptibility tests and the phenotypic method.

Total DNA extraction. Total DNA of MRSA isolates were extracted by the boiling method described by Zhang et al. (9).

Detection of resistance and the pvl gene by PCR.

PCR amplification of the *msrA/B*, *ermA*, *ermB*, *ermC*, *mecA* and *pvl* genes were done with primers listed in Table 1 as described previously (3, 4, 6, 10-13). The PCR products were observed by electrophoresis on 1% agarose gels (SinaClon, Co, Iran). This was followed by DNA Green Viewer (Pars Tous Biotechnology, Co, Iran) staining and analysis.

Detection of SCC*mec* types by multiplex PCR. Two different multiplex PCR methods, described by Oliveira et al. and Boye et al., were used for SCC*mec* typing of MRSA isolates (4, 5).

RESULTS

Of the 81 clinical samples collected from different patients, 8 isolates were identified as *S. aureus* using standard microbiological tests and amplification of the *nuc* gene. Of the 8 *S. aureus*, two isolates

Table 1. The list of oligonucleotide primers were used in this study.

Targetgene	Primer sequence (5'-3')	Annealing temperature (°C)	Product size (bp)	Reference
пис	F-GCGATTGATGGTGATACGGTT	60	279	(6)
	R-AGCCAAGCCTTGACGAACTAAAGC			
mecA	F-TCCAGATTACAACTTCACCAGG	56	162	(4)
	R-CCACTTCATATCTTGTAACG			
ermA	F-TATCTTATCGTTGAGAAGGGATT	56.5	139	(10)
	R-CTACACTTGGCTTAGGATGAAA			
ermB	F-CTATCTGATTGTTGAAGAAGGATT	55.5	142	(3)
	R-GTTTACTCTTGGTTTAGGATGAAA			
ermC	F-AATCGTCAATTCCTGCATGT	55.5	297	(11)
	R-TAATCGTGGAATACGGGTTTG			
msrA	F-GCAAATGGTGTAGGTAAGACAACT	56.5	402	(12)
	R-ATCATCATGTGATGTAAACAAAAT			
pvl	F-ATCATTAGGTAAAATGTCTGGACATGATCCA	55	433	(13)
	R-GCATCAASTGTATTGGATAGCAAAAGC			

were from the surgical external ocular infection of two males (43 and 82-years old) with diabetes mellitus. One isolate was from the surgical external ocular infection of a 73-year-old female with diabetes mellitus, four isolates were from external ocular infection from 27, 47, 48 and 64-years-old males and one of the isolates was from external ocular infection a 21-year-old female. All isolates were susceptible to linezolid and vancomycin. Of the 8 S. aureus, 5 isolates were methicillin-sensitive Staphylococcus aureus (MSSA) and were susceptible to amikacin, gentamicin, ciprofloxacin, clindamycin, erythromycin, trimethoprim/sulfamethoxazole, and tetracycline. The pvl, mecA, ermA, ermB, ermC and msrA/B genes were not detected in MSSA isolates. Three (37.5%) S. aureus isolates were resistant to cefoxitin and considered as MRSA. The isolates were not inducible clindamycin resistance. The ermC and pvl genes were only detected in a MRSA isolate obtained from external ocular infection of a 43-year-old male with diabetes mellitus. This isolate belonged to SCCmec IV and was resistant to trimethoprim/sulfamethoxazole, amikacin, gentamicin, ciprofloxacin, clindamycin, erythromycin, and tetracycline. Two MRSA isolates were from external ocular infection of 47 and 63-years-old males were not typeable with SCCmec typing method, were susceptible to amikacin, gentamicin, ciprofloxacin, clindamycin, erythromycin, tetracycline, and trimethoprim/sulfamethoxazole.

DISCUSSION

According to literature, *S. aureus* is the most important Gram-positive bacterium in ocular infections (1, 2). A wide range of resistance genes in *S. aureus* isolates has been reported from different samples in previous studies (1-3). Epidemiological investigations revealed that ocular infections by MRSA are increasing worldwide (14). Also, many studies in the ocular infection literature have shown that MRSA isolates are increasing in local settings (1, 2). As well, data suggested CA-MRSA isolates are the major threat in the ophthalmic settings and ocular infections (1, 2).

According to epidemiological evidence, prevalence of MRSA isolates in eye infections is 34% to 53% in different countries such as the United States, India and Taiwan (15, 16). The most important challenge in dealing with MRSA bacteria is to reduce the antibiotic choices in empirical therapy or prophylaxis because these isolates are commonly multi-drug resistant (MDR) (1). Diabetic patients are at a higher risk of getting infections than non-diabetics and ocular infection usually occurs in diabetic patients after eye surgery (2). In our study, all three MRSA bacteria were obtained from diabetic patients. Several studies, reported spread of MRSA isolates among ocular infections (1, 2, 16, 17). In a study in 2010 by Khan et al., MRSA isolates, belonging to SCCmec II and III, were reported in patients with ocular infections (18). MRSA isolates with SCCmec types I to III were

considered as HA-MRSA and MRSA isolates with SCCmec type IV were considered as CA-MRSA (19). Detection of SCCmec IV and pvl genes in a MRSA isolate in our study revealed this isolate closely related to CA-MRSA (2, 3, 20). CA-MRSA isolates usually harbor panton-valentine leukocidin (PVL), therefore, ocular infections caused by CA-MRSA leads to more damage to the eye (16). In the present study, two MRSA isolates were not typeable in SC-Cmec typing. These results showed that other types of SCCmec may be spreading among MRSA isolates from ocular infections in our region.

In conclusion, MRSA isolates are commonly multi-drug resistant, therefore, antibacterial susceptibility test results should be considered in treatment of eye infections. On the other hand, CA-MRSA are usually PVL positive which increases eye damage, hence, genetic characteristics and molecular typing of MRSA isolates are very useful for detection and infection control.

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REFERENCES

- Shanmuganathan VA, Armstrong, Buller A, Tullo AB. External ocular infections due to methicillin-resistant *Staphylococcus aureus* (MRSA). *Eye (Lond)* 2005;19:284-291.
- 2. Chuang CC, Hsiao CH, Tan HY, Ma DH-K, Lin KK, Chang CJ, et al. *Staphylococcus aureus* ocular infection: methicillin-resistance, clinical features, and antibiotic susceptibilities. *PLoS One* 2012;7:e42437.
- 3. Chaieb K, Zmantar T, Chehab O, Bouchami O, Ben Hasen A, Mahdouani K, et al. Antibiotic resistance

- genes detected by multiplex PCR assays in *Staphylococcus epidermidis* strains isolated from dialysis fluid and needles in a dialysis service. *Jpn J Infect Dis* 2007;60:183-187.
- Oliveira DC, de Lencastre H. Multiplex PCR strategy for rapid identification of structural types and variants of the mec element in methicillin-resistant Staphylococcus aureus. Antimicrob Agents Chemother 2002;46:2155-2161.
- Boye K, Bartels MD, Andersen IS, Mølle JA, Westh H. A new multiplex PCR for easy screening of methicillin-resistant *Staphylococcus aureus* SCC*mec* types I–V. *Clin Microbiol Infect* 2007;13:725-727.
- Sadeghi J, Mansouri S. Molecular characterization and antibiotic resistance of clinical isolates of methicillin-resistant *Staphylococcus aureus* obtained from Southeast of Iran (Kerman). *APMIS* 2014;122:405-411.
- Sedaghat H, Nasr Esfahani B, Mobasherizadeh S, SallariJazi A, Halaji M, Sadeghi P, et al. Phenotypic and genotypic characterization of macrolide resistance among *Staphylococcus aureus* isolates in Isfahan, Iran. *Iran J Microbiol* 2017;9:264-270.
- 8. Performance standards for antimicrobial susceptibility testing (CLSI). 27th ed. CLSI supplement M100.Wayne, PA: Clinical and Laboratory Standards Institute; 2017.
- Zhang K, McClure JA, Elsayed S, Louie T, Conly JM. Novel multiplex PCR assay for characterization and concomitant subtyping of staphylococcal cassette chromosome mec types I to V in methicillin resistant Staphylococcus aureus. J Clin Microbiol 2005;43:5026-5033.
- 10. Zmantar T, Kouidhi B, Miladi H, Bakhrouf A. Detection of macrolide and disinfectant resistance genes in clinical *Staphylococcus aureus* and coagulase-negative staphylococci. *BMC Res Notes* 2011;4:453.
- Strommenger B, Kettlitz C, Werner G, Witte W. Multiplex PCR assay for simultaneous detection of nine clinically relevant antibiotic resistance genes in *Staphylo*coccus aureus. J Clin Microbiol 2003;41:4089-4094.
- Singh KV, Malathum K, Murray BE. Disruption of an *Enterococcus faecium* species-specific gene, a homologue of acquired macrolide resistance genes of staphylococci, is associated with an increase in macrolide susceptibility. *Antimicrob Agents Chemother* 2001;45:263-266.
- Lina G, Piémont Y, Godail-Gamot F, Bes M, Peter MO, Gauduchon V, et al. Involvement of Panton-valentine leukocidin-producing *Staphylococcus aureus* in primary skin infections and pneumonia. *Clin Infect Dis* 1999;29:1128-1132.
- 14. Havaei SA, Halaji M, Vidovic SR. Dillon J, Karbalaei M, et al. Prevalence and genotyping of methicillin-resistant and susceptible *Staphylococcus aureus* strains isolated from patients in a university hospital, Isfahan, Iran. *Jundishapur J Microbiol* 2017;10:e13571.

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- Vola ME, Moriyama AS, Lisboa R, Vola MM, Hirai FE, Bispo PJ, et al. Prevalence and antibiotic susceptibility of methicillin-resistant *Staphylococcus aureus* in ocular infections. *Arq Bras Oftalmol* 2013;76:350-353.
- 16. Hsiao CH, Ong SJ, Chuang CC, Ma DHK, Huang YH. A comparison of clinical features between community-associated and healthcare-associated methicillin-resistant *Staphylococcus aureus* Keratitis. *J Ophthalmol* 2015;2015:923941.
- 17. Freidlin J, Acharya N, Lietman TM, Cevallos V, Whitcher JP, Margolis TP. Spectrum of eye disease

- caused by methicillin-resistant *Staphylococcus aureus*. *Am J Ophthalmol* 2007;144:313-315.
- Khan MA, Ahmad S, Banu N. Molecular characterisation of methicillin-resistant *Staphylococcus aureus* (MRSA) from keratitis patients: a microbiological analysis. *Br J Ophthalmol* 2010;94:994-998.
- 19. Loomba PS, Taneja J, Mishra B. Methicillin and vancomycin resistant *S. aureus* in hospitalized patients. *J Glob Infect Dis* 2010;2:275-283.
- 20. Deurenberg RH, Stobberingh EE. The evolution of *Staphylococcus aureus. Infect Genet Evol* 2008; 8:747-763.