

Frequency of *Streptococcus pneumoniae* infection in patients with suspected meningitis in Imam Reza Hospital of Kermanshah in the west of Iran

Kamal Ahmadi¹, Alisha Akya^{2*}, Bizhan Numanpour^{1,3}, Afsaneh Salimi⁴, Asad Veisi - Raigani⁴

Student Research Committee, Faculty of Medicine, Kermanshah University of Medical Sciences, Kermanshah, Iran. 2 Nosocomial Infection Research Centre, School of Medicine, Kermanshah University of Medical Sciences, Kermanshah, Iran. ³Department of Microbiology, School of Medicine, Kermanshah University of Medical Sciences, Kermanshah, Iran. ⁴Department of Biochemistry, School of Medicine, Kermanshah University of Medical Sciences, Kermanshah, Iran.

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ABSTRACT

Background and Objectives: Streptococcus pneumoniae is the main causative agent of acute bacterial meningitis in the world. This study aimed to detect pneumoniae infection in cerebrospinal fluid (CSF) samples of patients with suspected meningitis.

Materials and Methods: In this study, 200 CSF samples in patients with suspected meningitis and with negative bacterial cultures were tested. Demographic data of patients and the laboratory analysis of CSF samples were also collected in a questionnaire. Pneumococcal capsular gene spn9802 was examined by real-time PCR technique.

Results: Of 200 CSF samples from patients with the average age of 32.1±25.3 year old, 20 were positive for S. pneumoniae in patients with the average age of 35.05±24.6. The biochemical and cytological analysis of positive samples showed significant changes, including, 39.7 mg/DL protein, 46.2 mg/DL glucose and 1173 white blood cells per microliter of CSF on average. Conclusion: The results of this study showed the significant pneumococcal infection in culture negative CSF samples. The majority of this infection occurred in middle-aged patients and with a higher incidence rate in the winter. It seems that the traditional methods of cultivation are not sensitive enough to detect this bacterium in CSF .Alternatively, the molecular techniques such a real-time seem to be accurate, sensitive and rapid for the detection of this agent in CSF. The cytological and biochemical findings of CSF can provide valuable clues in the diagnosis of bacterial meningitis.

Keywords: Culture-negative meningitis, real-time PCR, Streptococcus pneumoniae

INTRODUCTION

of illnesses, including pneumonia, sinusitis, otitis

Streptococcus pneumoniae can cause a range

*Corresponding author: Dr. Alisha Akya Address: Department of Microbiology, Faculty of Medicine, Kermanshah University of Medical Sciences, Shirudi Boulevard, Kermanshah, 6714869914, Iran.

Tel: +98 8314262252 Fax: +98 831 4262252 E-mail: akya359@yahoo.com media, bacteremia, cellulitis, and acute bacterial meningitis (1, 2). Every year a large number of people, particularly in developing countries, lose their lives from different diseases caused by this bacterium (3,4). S. pneumoniae is the leading cause of acute bacterial meningitis in adults and young people (5). Meningitis caused by this bacterium has a higher rate of mortality than any other bacteria that cause this disease (16 to 37%) (6). Permanent neurological disabilities occur in 30 to 52% of surviving patients (7,8). Traditional methods used in the diagnosis of meningitis include microscopy, bacterial culture, cytology and biochemical analysis

of CSF, and serological methods (1,2,4,9). Direct examination of smears prepared from CSF is a quick method to make a clinical decision but needs at least 10⁴ to 10⁵ bacteria per mL of CSF to get a positive result. This factor reduces the sensitivity of this method. As afastidious organism, *S. pneumoniae* grows very difficult using conventional culture media and requires at least 24-48 hours time (10,11). Due to the risk of death or permanent neurological sequels, patients with suspected bacterial meningitis usually receive antibiotics in the first medical visit prior to sampling. This can kill the bacteria in sterile fluids of the body within 2-4 hours (12).

Consequently, it might result in a negative culture of bacteria and thus reduces the sensitivity of this method to approximately 30% in medical diagnostic laboratories (13). Therefore, if the conventional bacteriogy is used for diagnosis, as it occurs in many medical laboratoirs in Iran, the prevalence of pneumococcal meningitis is underestimated. Alternative methods based on DNA amplification using distinct target genes have been developed for the detection and rapid diagnosis of pneumococcal meningitis (11,14). Given the advantages, including the reducing of time, the elimination of post PCR processing and quantitative analysis, real-time PCR method has been updated for this purpose (15). Furthermore, the rate of possible contamination of samples is lower in real-time PCR than conventional PCR, making it more sensitive and specific technique for diagnosis (15-17). The purpose of this study was to evaluate the quantitative real time PCR assay for detection of S. pneumoniae from CSF specimens of patients suspected with meningitis.

MATERIALS AND METHODS

Sampling. A total of 200 culture-negative CSF samples from patients suspected with meningitis,

according to an infectious disease specialist examination were investigated in this study. Samples were collected during 2011 to 2013 from patients in the main hospital (Imam Reza) in Kermanshah city located in the west of Iran. The samples were quickly transported in an ice box under sterile conditions to the microbiology laboratory and kept at -70 °C until DNA was extracted (2). CSF samples with volume less than 50 μ l and samples with positive bacterial cultures were exluded from the study.

Real-time PCR assay. The 100 µl of CSF samples were used for DNA extraction by CinnaPure DNA kit (SinaClon, Iran) according to manufacturer's instructions. The concentration of extracting DNA (OD 260/280NM) was determined by spectrophotometry and gel electrophoresis and the specificity and characteristics of the DNA were determined using the internal control. Extracted DNA was kept at minus 20°C until used in real-time PCR assay. The following primers were used to amplify a part of spn9802 gene (18) of S. pneumoniae, F: (AGTCGTCCAAGGTAAACAAGTCT) and (ACCAACTCGACCACCTCTTT). Real-time PCR reaction was prepared in a final volume of 20µl, using 6 µl of Master mix Eva Green, 0.5 µl of each primer, 9µl deionized distilled water and 4µl of DNA template. The real-time PCR apparatus was Rotor-Gene 6000 (Corbett Research, Australia). The initial denaturing temperature of 95 °C was used for 10 min followed by 40 cycles of 95°C for 30 s, and 60°C for 60 seconds.

Five serial dilutions of standard bacterial DNA contained the 10^1 to 10^5 copies per mL of bacterial genome were prepared as positive control reactions in each run of real-time PCR. At each stage, based on E (efficiency) equals to 1.02 ± 0.05 , and the amount of R_2 (Coefficient of Determination) and R (Regression) which was lower than 0.975, the test was repeated.

Table 1. The results of serial dilutions of *S.pneumoniae*(PTCC1240)

No.	Std*	CFU/mL	CT	Bacterial Count(CFU/mL)
1	S ₁	10 5	19.42	1.5e+5
2	\mathbf{S}_2	10 4	22.64	1.7e+4
3	S_3	10 ³	25.35	2.5e+3
4	S_4	10 ²	28.03	5.5e+2
5	S_5	10 1	32.11	7.6e+1

Std*=Standard

Statistical analysis. All collected data were statistically analyzed using SPSS version 18 and the comparison of data was done using t-test.

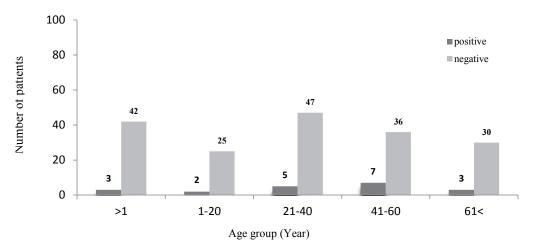


Fig. 1. The age distribution of patients with positive and negative cases.

Specificity of reaction. Genome of various bacteria including Staphylococcus aureus (PTCC 1189), Enterococcus faecalis (PTCC 1237), Streptococcus epidermidis (PTCC 1435), Streptococcus agalactiae (PTCC 1768), Streptococcus pyogenes (PTCC 1447); Neisseria meningitidis (ATCC 13090); Escherichia coli (PTCC 1395), Klebsiella pneumoniae (PTCC 1053), Pseudomonas aeruginosa (PTCC 1310), Haemophilus influenzae (PTCC 1766), Salmonella typhi (PTCC 1609); and gram-positive bacilli: Listeria menocytogenes (PTCC 1297) and human DNA were extracted and used as controls in real-time reactions.

Determination of sensitivity of reaction. To determine the sensitivity of the method, the serial dilutions of 10¹ to 10⁵ copies per mL of *S. pneumoniae* (PTCC1240) genome were prepared. These serial dilutions were used for making the standard curve of real-time PCR (Table 1).

RESULTS

Of 200 cases, 99 (%49) and 101 (%51) were males and females, respectively. Their age ranged from one day to 91 year old with an average age of 35.05± 24.6 years. In total, 20 cases of pneumococcal infections were detected in CSF samples (Table 2 and Fig. 2). Of these, 11 and 9 were males and females, respectively, with an average age of 35.5 years old (Fig. 1). Seven cases occurred in winter that was slightly higher than other seasons. The average number of bacterial genomes in CSF samples was 2.6×10⁴copies/ml. The statistical results of the comparison of positive and negative samples are presented in Table 3.

DISCUSSION

Despite great advances in medical knowledge, bacterial meningitis remains as one of the infectious disease with high mortality rate. S. pneumoniae is the major cause of bacterial meningitis in all ages in many countries, although with different rates of prevalence. Our results showed 10% of culture-negative CSF was positive for S. pneumoniae infection. That indicates the significant rate of pneumococcal meningitis among undiagnosed CSF samples. In many cases, CSF samples are not routinely tested by molecular methods to detect this bacterium (2,9,13,14,19). Studies in different countries using PCR and realtime PCR methods, have reported the positive cases of pneumococcal infection in culture-negative CSF samples. In Brazil, of 428 CSF samples tested, 70 cases were positive (16.35%), for S. pneumoniae while in the United States, 5 samples out of 11 (45.45%) and in England 3 out of 7 (42.85%) samples gave positive results for this organism as causative of infection(5,19,20). Because of better primary examination and evaluation by doctors, it seems that in developed countries more cases can be detected. Coveresly, in India 2 out of 43 samples (4.65%) were positive for S. pneumoniae in cases suspected with meningitis (14).

The conventional methods for identification of bacterial meningitis need to have at least 10⁴ to 10⁶ live bacteria per mL of CSF that is hard to occur because of antibiotic consumption before sampling (10,11). Our results indicated the range of bacterial genome copy was from 50 to 150000 per mL of CSF. This is mainly less than limit range for conventional culture methods and explained, at least in part, the

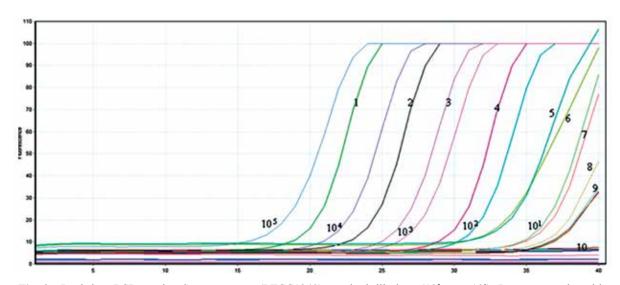


Fig. 2. Real-time PCR results. *S. pneumoniae* (PTCC1240) standard dilutions (10⁵ up to10¹), Pneumococcal positive Samples (number 1 to 6), Negative Samples (7 to 9), Negative control (10).

Table 2. Biochemical and cytological characteristics of positive samples.

Gender		Protein mg/dL	Glucose mg/dL	WBC	Differential counts			Bacterial count
	Age				Seg. %	Lym. %	CT	(copies/mL)
M	75	27	91	25	75	25	23.07	(2.7e+3)
M	73	40	41	700	70	30	20.81	(9.4e+4)
M	69	16	70	8000	90	10	19.45	(1.4e+5)
M	53	37	20	160	15	85	23.94	(1.5e+3)
M	50	65	55	8000	90	10	19.64	(1.5e+5)
M	47	32	52	200	50	50	24.27	(1.4e+3)
M	32	20	42	3200	80	20	22.83	(1.3e+4)
M	26	90	35	110	75	25	21.91	(1.5e+4)
M	19	25	36	140	20	80	26.02	(1.3e+3)
M	C*	40	20	50	70	30	30.51	(8.1e+2)
M	C*	42	66	100	90	10	20.97	(6.5e+4)
F	60	20	47	100	40	60	26.01	(1.1e+3)
F	53	65	22	900	70	30	22.64	(1.4e+4)
F	42	100	39	1350	85	15	22.59	(9.8e+3)
F	35	18	82	53	85	15	24.28	(1.4e+3)
F	24	20	55	50	82	18	24.37	(1.3e+3)
F	23	28	55	50	80	20	24.76	1.3e+3
F	19	56	23	200	70	30	25.66	(2.5e+3)
F	C*	18	43	40	60	40	33.03	(5e+1)
F	C*	40	41	40	75	25	25.48	(1.1e+3)

C*= Children less than a year old F= Female M=Male CT=Cycle threshold

Table 3. Comparison analysis of positive and negative samples.

Characteristic	Negative samples (180)	Positive samples (20)	p-value	
Average age	36.23	35.05	0.24	
Protein	28.27	39.95	0.001	
Glucose	62.26	46.75	0.008	
WBC count	203.34	1174.40	0.041	

lower sensitivity of conventional methods.

In bacterial meningitis, biochemical and cytological characteristics of CSF can provide valuable clues for diagnosis. Among these characteristics, increased in the number of white blood cells (WBC), increase in protein level and the decrease of glucose level in CSF are helpful clues. However, these factors cannot unequivocally differentiate viral from bacterial cases (21,22).

In our study, a significant increase in the number of WBC and protein level and decrease in glucose level was observed, which is consistent with the medical context (26). In terms of age distribution of patients, the results of other studies indicated that the S. pneumoniae is the main cause of bacterial meningitis in elderly people with the age range 40 to 60 years. which is in agreement with our findings (23,24). In terms of the incidence of pneumococcal meningitis in men and women, our results showed the slightly higher rate of the positive cases among men, which is consistent with other reports (25,26). The incidence of bacterial meningitis occurs mostly in winter (26-28) and that is similar to our results. However, because of a small number of positive cases, seasonal pattern was not statistically approved.

CONCLUSION

The results of this study showed the significant pneumococcal infection rate in culture - negative CSF samples in Kermanshah. The majority of these infections occurred in middle-aged patients and with the slightly higher incidence rate in the winter. It seems that the traditional methods of cultivation are not sensitive enough to detect this bacterium in CSF. Alternatively, real-time PCR seems to be accurate, sensitive and rapid for the detection of this agent in CSF. The cytological and biochemical findings of CSF analysis, including the increase in the number of WBC and protein level and decrease in the glucose level can provide valuable clues for the diagnosis of bacterial meningitis.

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REFERENCE

- Rintamäki S, Saukkoriipi A, Salo P, Takala A, Leinonen M. Detection of *Streptococcus pneumoniae* DNA by using polymerase chain reaction and microwell hybridizationwith Europium-labelled probes. *J Microbiol Methods* 2002; 50: 313-318.
- Greiner O, Day PJR, Bosshard PP, Imeri F, Altwegg M, Nadal D. Quantitative detection of *Streptococcus* pneumoniae in nasopharyngeal secretions by real-time PCR. J Clin Microbiol 2001; 39: 3129-3134.
- 3. Kim DW, Kilgore PE, Kim EJ, Kim SA, Anh DD, Dong BQ, et al. The Enhanced *Pneumococcal* LAMP Assay: A clinical tool for the diagnosis of meningitis due to *Streptococcus pneumoniae*. *PLoS One* 2012; 7: 1-8.
- Williams TM, Loman NJ, Ebruke C, Musher DM, Adegbola RA, Pallen MJ, et al. Genome analysis of a highly virulent serotype 1 strain of *Streptococcus* pneumoniae from West Africa. PLoS One2012; 7(10) :1-10.
- Cherian T, Lalitha MK, Manonaran A, Thomas K, Yolken RH, Steinhoff MC. PCR-enzyme immunoassay for detection of *Streptococcus pneumoniae* DNA in cerebrospinal fluid samples from patients with culturenegative meningitis. *J Clin Microbiol* 1998; 36: 3605-3608.
- 6. Weisfelt M, Beek DVD, Spanjaard L, Reitsma JB, Gans JD. Clinical features, complications, and outcome in adults with pneumococcal meningitis: a prospective case series. *Lancet Neurol* 2006; 5: 123-129.
- Chavez-Bueno S, McCracken GH Jr. Bacterial meningitis in children. *Pediatr Clin North Am* 2005; 52: 795-810.
- 8. Kastenbauer S, Pfister H-W. *Pneumococcal* meningitis in adults: spectrum of complications and prognostic factors in a series of 87 cases. *J Brain Dis* 2003; 126: 1015-125.
- 9. Tunkel AR, Hartman BJ, Kaplan SL, KaufmanBA,

- Roos KL, Scheld MW, et al. Practice guidelines for the management of bacterial meningitis. *Clin Infect Dis* 2004;39:1267-84.
- 10. Failace L, Wagner M, Chesky M, Scalco R, Jobim LF. Simultaneous detection of *Neisseria meningitidis*, *Haemophilusinfluenza and Streptococcussp*. by polymerase chain reaction for the diagnosis of bacterial meningitis. *Arg Neuropsiquiatr* 2005; 63: 920-924.
- GhotaslouR, Farajnia S, Yeganeh F, Abdoli-Oskouei S, Ahangarzadeh-Rezaee M, Barzegar M. Detection of acute childhood meningitis by PCR, culture and agglutination tests in Tabriz, Iran. *Acta Medica Iranica* 2012; 50: 192-196.
- 12. Kanegaye JT, Soliemanzadeh P, Bradley JS. Lumbar puncture in pediatric bacterial meningitis: defining the time interval for recovery of cerebrospinal fluid pathogens after parenteral antibiotic pretreatment. *JAMA Pediatr* 2001; 108: 1169-1174.
- Corless CE, Guiver M, Borrow R, Edwards-Jones V, Fox AJ, Kaczmarski EB. Simultaneous detection of Neisseria meningitidis, Haemophilus influenzae, and Streptococcus pneumoniae in suspected cases of meningitis and septicemia using real-time PCR. J Clin Microbiol 2001; 39: 1553-1558.
- PanditL, Kumar S, Karunasagar I, Karunasagar I. Diagnosis of partially treated culture-negative bacterial meningitis using 16S rRNA universalprimers and restriction endonuclease digestion. *J Med Microbio* 2005; 54: 539-542.
- Espy MJ, Uni jR, Sloan LM, Buckwalter SP, Jones MF, Vetter EA, et al. Real-Time PCR in Clinical Microbiology: Applications for Routine Laboratory Testing. J ClinMicrobiol Reviews 2006; 19: 165-256.
- 16. Saukkoriipi A, Palmu A, Kilpi T, Leinonen M. Real-time quantitative PCR for the detection of *Streptococcus* pneumoniae in the middle ear fluid of children with acute otitis media. Mol Cell Probes 2002; 16: 385-390.
- 17. Bryant PA, Li HY, Zaia A, Griffith J, Hogg G, Curtis N, et al. Prospective study of a real-timePCR that is highly sensitive, specific, and clinically useful for diagnosis of meningococcal disease in children. J ClinMicrobiol 2004; 42: 2919-2925.

- 18. Suzuki N, Seki M, Nakano Y, Kiyoura Y, Maeno M, Yamashita Y. Discrimination of *Streptococcus pneumoniae* from viridans group*streptococci* by genomic subtractive hybridization. *J Clin Microbiol* 2005; 43: 4528-4534.
- 19. Sacchi CT, Fukasawa LO, Goncalves MG, Salgado MM, Shutt KA, Carvalhanas TR, et al. Incorporation of real-time PCR into routine public health surveillance of culture negative bacterial meningitis in Sa o Paulo, Brazil. *PLoS One*2011; 6: 1-8.
- Kearns AM, Freeman R, Murphy OM, Seiders PR, Steward M, Wheeler J. Rapid PCR-based detection of Streptococcus pneumoniae DNA in cerebrospinal fluid. J Clin Microbiol 1999; 37: 3434.
- Reilly BM, Evans AT.Translating clinical research into clinical practice: Impact of using prediction rules to make decisions. *Ann Intern Med* 2006; 144: 201-209.
- Negrini B, Kelleher KJ, Wald ER.Cerebrospinal fluid findings in aseptic versus bacterial meningitis. *JAMA Pediatr* 2000;105: 316-319.
- Ataee RA, Mehrabi-Tavana A, Izadi A, Hosseini SMJ, Ataee MH. Bacterial meningitis: a new risk factor. *JRMS* 2011;16: 207-210.
- Brindusa T, Klara B, Tilea I.Incidence, epidemiology and etiology of acute meningitis in Mures County–an observational study. ARS Medica Tomitana 2012; 3: 116-125
- Mosavi-Jarrahi A, Abdolreza Esteghamati A, Asgari F, Heidarnia M, Mousavi-Jarrahi Y, Goya M. Temporal analysis of the incidence of meningitis in the Tehranmetropolitan area, 1999-2005. *Population Health Metrics* 2009; 7: 1-7.
- Bahador M, Amini M, Bahador M. Common cause and cerebrospinal fluid changes of acute bacterial meningitis. *IJP* 2009; 4:75-79.
- 27. Thomas JD, Hatcher CP, Satterfield DA, Theodore MJ, Bach MC, Linscott KB, et al. SodC-Based Real-Time PCR for Detection of *Neisseria Meningitidis*. *PLoS One*2011;6: 1-8.
- 28. Mandell, Douglas, Bennett's (2010). Principles and practice of infectious diseases. 7th edChurchill Livingstone Elsevier.